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Indoor potted plants of Marantaceae and Pteridophytes for purification of formaldehyde polluted air

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Abstract: Ten plants from Marantaceae families and ten plants from Pteridophytes were tested for their abilities of removing formaldehyde(FDH) in the air. Each of the plants was placed for seven days in a 1.0 m×1.0 m×0.8 m glass box filled with FDH with the initial concentration of 15 mg/m³. Some plants such as *Neottopteris nidus* Volulum, *Calathea lubbersiana* showed the top resistant ability to FDH; some plants such as *Calathea ornata*, *Calathea setosa*, *Calathea freddy*, and *Calathea roseo-picta* showed medium resistant ability to FDH; other species such as *Neottopteris nidus*, *Pteris fauriei*, *Pteris ensi formis* cv. Victoriae, *Pteris cretica* cv. Albolineata, *Nephrolepis cordifolia*, *Cyclosorus parasiticus*, *Blechnum orientale*, *Maranta bicolor*, and *Calathea zebrina* showed no resistance to FDH. The absorption of plants for FDH in the glass chamber was found especially apparently different during the first three days. The fastest purification rates of FDH were found in species such as *C. zebrina*, *Microsorium punctatum*, and the slowest were found in species such as *C. parasiticus*, *P. ensi formis* cv. Victoriae, *N. nidus* cv. Volulum, and *C. setosa*. In conclusion, 8 species of the potted plants which could be recommendable to be used for FDH purification were *N. nidus*, *Calathea rotundi folia*, *P. cretica* cv. Albolineata, *C. ornata*, *Platyserium bifurcatum*, *N. nidus* cv. Volulum, *C. roseo-picta*, and *C. freddy*, because these plants have high absorption ability to FDH and have less damage.

Key words: formaldehyde; indoor potted plants; Marantaceae; Pteridophytes; purification

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竹芋科和蕨类室内盆栽植物对 甲醛净化作用的研究

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摘要: 以竹芋科和蕨类室内盆栽植物各 10 种为试验材料, 分别置于体积为 1.0 m×1.0 m×0.8 m 密封玻璃箱内, 甲醛起始浓度均设置为 15 mg/m³, 连续观察 7 d。结果表明: 卷叶巢蕨(*Neottopteris nidus* cv. Volulum)、矩叶肖竹芋(*Calathea lubbersiana*)对甲醛抗性最强(I级); 银线竹芋(*C. ornata*)、银羽斑竹芋(*C. setosa*)、翠叶竹芋(*C. freddy*)和彩虹竹芋(*C. roseo-picta*)抗性较强(II级); 巢蕨(*N. nidus*)、傅氏凤尾蕨(*Pteris fauriei*)、银脉凤尾蕨(*P. ensi formis* cv. Victoriae)、银心大叶凤尾蕨(*P. cretica* cv. Albolineata)、肾蕨(*Nephrolepis cordifolia*)、华南毛蕨(*Cyclosorus parasiticus*)、乌毛蕨(*Blechnum orientale*)、花叶竹芋(*Maranta bicolor*)和天鹅绒竹芋(*C. zebrina*)抗性最差(IV级)。甲醛处理后, 密封玻璃箱内甲醛浓度均呈递减变化, 递减最快都集中在试验后 1~3 d 之间。吸收甲醛最快的植物是天鹅绒竹芋和星蕨(*Microsorium punctatum*), 最慢的是华南毛蕨、银脉凤尾蕨、卷叶巢蕨和

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银羽斑竹芋。对甲醛处理产生伤害反应少或较少,而吸收能力强的前 8 种植物是:巢蕨、青苹果竹芋(*C. rotundifolia*)、银心大叶凤尾蕨、银线竹芋、二歧鹿角蕨(*Platycerium bifurcatum*)、卷叶巢蕨、彩虹竹芋和翠叶竹芋,可作为甲醛净化专用植物应用推广。

关键词: 甲醛; 盆栽植物; 竹芋科; 蕨类; 净化作用

Increasing uses of resins and solvents such as formaldehyde (FDH) in construction and decoration materials have caused severe pollution of indoor air. FDH is a kind of colorless chemical with a strong pungent odor, especially steadily releasing from indoor decoration materials for three to fifteen years. FDH like other chemicals such as benzene causes serious hazards to human health and is classified as the first class of human carcinogens by International Agency for Research on Cancer (IARC) because it is capable of inducing cancers and tumors.

Interest in the indoor air quality has become an earnest issue in China since there is a serious problem of excessive use of FDH in housing construction, reformation and decoration. Yu & Tang (2005) monitored more than fifty new decorated bedrooms and found that the pollutants such as FDH, benzene and other volatile organic compounds exceeded the limited standards by 36.8%, 38.4% and 12%, respectively. The adoption of energy-saving proposals to reduce the release of indoor pollutants has led inefficiency in improvement of indoor air quality and consequently makes potential health hazards remaining. It is one of today's hottest research subjects how to use indoor plants to absorb and remove the indoor air pollutants. The early research was carried out by Wolverton of National Aeronautics and Space Administration (NASA) and the research group listed the top ten plants which were effective in clearing and eliminating odor, purifying air, and absorbing and decomposing FDH, benzene and other pollutants (Wolverton, 1997; Wolverton *et al.*, 1985). These plants include spider plant (*Chlorophytum comosum*), English ivy (*Hedera nepalensis* var. *sinensis*), Aloe vera, *Dracaena sanderiana*, *D. marginata*, *D. fragrans* var. *Massangeana*, mother-in-law's tongue (*Sansevieria trifasciata*), *S. trifasciata*, *Gerbera jamesonii*, Chinese evergreen (*Aglaonema* spp.), peace lily (*Spathiphyllum floribundum* cv. *Clevelandii*), and

Pritchardia gaudichaudii. Recent years, many researchers and scientists in China focused on selection of plant species that are effective in absorbing or removing FDH from indoor air (Li, 2006; Zhou *et al.*, 2006; Huang *et al.*, 2008; Cao *et al.*, 2009; Xiong & Su, 2009; Wu 2006). Plants such as *Monstera deliciosa*, *Ficus elastic*, *Chlorophytum comosum*, and *Opuntia stricta* have proved capable of absorbing FDH in the air. However, effective species are still limited. Noticeably, it is still difficult to eliminate FDH, benzene and other indoor air pollutants in low concentrations. The previous studies showed that many potted plants had more or less abilities of absorbing and decomposing FDH and benzene. Moreover, the pots, culture media and microorganisms in the media also showed ability of absorbing FDH to some extent. However, the purification effects of the tested plants, such as *Chlorophytum comosum* and *Scindapsus aureun*, were inconsistent in different researches. Although ten species of the plants such as *Scindapsus aureun*, *Asparagus setaceus*, *Sansevieria trifasciata* cv. *Hahnii*, *Chlorophytum comosum*, *Aglaonema commutatum* cv. *White Rajah*, *Aglaonema commutatum* cv. *Red Narrow*, *Aglaonema commutatum* cv. *Traubii*, *Scindapsus pictus* cv. *Argyraeus*, *Gasteria gracilis*, *Philodendron sodiroi* cv. *Wendimbe* were recommended for FDH purification (Zhou *et al.*, 2011). But there is still much work to seek for more plants not only with good performance to remove air pollutants, but also with less or no damage to human beings. Therefore, in the present study, plants from the Marantaceae and Pteridophytes were tested for their abilities to absorb or remove indoor FDH and other pollutants as well as the resistance to FDH, in order to provide reference to the utilization of plants for indoor air purification.

1 Materials and Methods

Twenty potted plants from the Marantaceae

and Pteridophytes were used as experimental materials (Table 1).

Table 1 Twenty potted plants used from the Marantaceae and Pteridophytes

Marantaceae	Pteridophyte
<i>Calathea crocata</i>	<i>Blechnum orientale</i>
<i>Calathea freddy</i>	<i>Cyclosorus parasiticus</i>
<i>Calathea lubbersiana</i>	<i>Microsorium punctatum</i>
<i>Calathea makoyana</i>	<i>Neottopteris nidus</i>
<i>Calathea ornata</i>	<i>Neottopteris nidus</i> cv. Volulum
<i>Calathea roseo-picta</i>	<i>Nephrolepis cordifolia</i>
<i>Calathea rotundifolia</i>	<i>Platynerium bifurcatum</i>
<i>Calathea setosa</i>	<i>Pteris cretica</i> cv. Albolineata
<i>Calathea zebrina</i>	<i>Pteris ensiformis</i> cv. Victoriae
<i>Maranta bicolor</i>	<i>Pteris fauriei</i>

1.1 Experimental treatments

The tested potted plant was placed in a glass chamber with a wall 0.8 mm thick and inside volume of 0.8 m³ (1.0 m × 1.0 m × 0.8 m). Inside the chamber placed a small fan and a thermometer. The probe of formaldehyde (FDH) inspector (Ke Ernuo trading Co., Ltd. of Shenzhen) was inserted into the chamber through a hole of 1 cm diameter. The hole was removable and fitted with a rubber gasket and clamps to provide an airtight seal (Wolverton *et al.*, 1985). The initial concentration of formaldehyde was set up to 15 mg · m⁻³. The experimental temperature was controlled at (20 ± 1) °C. Three treatments were designed as followed: (1) plant potted in the medium was placed in the chamber with 15 mg · m⁻³ FDH; (2) the pot with medium but without plant was placed in the chamber with 15 mg · m⁻³ FDH; (3) the chamber only filled with 15 mg · m⁻³ FDH as control.

1.2 Measurement of chlorophyll

0.2 g fresh diachyma was obtained from the middle part of one leaf without venation before treatment and in the 7th day after being treated for measurement of chlorophyll, respectively. Leaf chlorophyll was extracted by 95% ethanol (Wang, 2006). The concentration of the extracted chlorophyll was measured at 665 nm and 649 nm wave length using a UV-Vis spectrophotometer (TU-1810 Model, General analysis instrument Co., Ltd. of Beijing). Concentrations of chlorophyll a (C_A), chlorophyll b (C_B) and the total chlorophyll (C_T) were calculated using equations as C_A =

13.7D₆₆₅ - 5.76D₆₄₉, C_B = 25.8D₆₄₉ - 7.6D₆₆₅, and C_T = C_A + C_B - 6.10D₆₆₅ + 20.04D₆₄₉, where, D₆₆₅ and D₆₄₉ were the optical density (OD) values of chlorophyll at 665 nm and 649 nm, respectively.

1.3 Measurement of leaf cell membrane permeability

The fresh diachyma was measured for cell membrane permeability before treatment and in the 7th day after being treated, respectively. The diachyma was rinsed 3 times with deionized water and the surface water was absorbed with filter paper. Then twenty leaf discs each in 0.5 cm diameter taken by hole puncher were immersed in 20 mL deionized water for three hours. The electrical conductivity (EC) (recorded as C₁) of the water after the leaf immersion was measured. Then the leaf sample was boiled for 15 min and the EC (C₂) was measured again after the water cooled down to the room temperature and was replenished to 20 mL with deionized water (Huang *et al.*, 1990). The leaf cell membrane permeability (L_c) was calculated as L_c (%) = 100 × (C₁/C₂).

1.4 Other measurements

The formaldehyde concentration in the chamber was measured every day after the experiment began. The fresh aboveground part taken from the plant was weighed as fresh mass. Then these materials were dried in the microwave oven for dry mass determination (Yu *et al.*, 2007). The damage of the tested plants in the formaldehyde air was scored from 0 to 4 degrees with the hurt increasing.

2 Results and Analysis

2.1 Plant response caused by formaldehyde

The ranking for hurt response of potted plants to formaldehyde (FDH) pollution was made shown in Table 2 according to (Zhou *et al.*, 2011). It was suggested that species like *N. nidus* 'Volulum' and *C. lubbersiana* showed more resistant ability to FDH damage and grouped into Grade 1. These plants such as *N. nidus*, *P. fauriei*, *P. ensiformis* cv. Victoriae, *P. cretica* cv. Albolineata, *N. cordifolia*, *C. parasiticus*, *B. orientale*, *M. bicolor*, and *C. zebrina* showed lower resistance to formaldehyde. The plants before (left in the frame) and after

Table 2 Damage response and grade for twenty potted plants after treated by FDH

Species	Response of potted plant	Grade
<i>N. nidus</i> cv. Volulum	A few spots on leaves	1
<i>C. lubbersiana</i>	Only 8 tiny spots on leaves	1
<i>C. ornata</i>	Only 1 etiolated lower leaf blade, more spots on other leaves, most leaves were normal	2
<i>C. setosa</i>	Only 2 etiolated lower leaf blades, a few spots on other leaves, most leaves were normal	2
<i>C. freddy</i>	Only 3 etiolated lower leaf blades, a few spots on other leaves, most leaves were normal	2
<i>C. roseo-picta</i>	Only 3 entirely brown lower leaf blades, more brown spots on other leaves, but stem in good condition	2
<i>P. bi furcatum</i>	Browning and rotten on half leaves. The plant would recover from the damage	3
<i>C. makoyana</i>	8 etiolated leaves, densely black or water stains spots on some leaves	3
<i>M. punctatum</i>	Black moldy on top leaf blades, normal damage on lower stem. The plant would recover from the damage	3
<i>C. crocata</i>	4 etiolated leaves, severe water stains spots on other leaves. The plant would recover from the damage	3
<i>C. rotundi folia</i>	5 etiolated leaves, many spots on other leaves, but stem in good condition. The plant would recover from the damage	3
<i>N. nidus</i>	Completely water stains spots and mould on most leaves	4
<i>P. fauriei</i>	Water stains spots on all leaves, plant dried	4
<i>P. ensiformis</i> cv. Victoriae	Water stains spots on all leaves, plant dried	4
<i>P. cretica</i> cv. Albolineata	Water stains on whole plant and died	4
<i>N. cordi folia</i>	Most leaves were dropped, water stains on whole plant and died	4
<i>C. parasiticus</i>	Water stains on whole plant and died	4
<i>B. orientale</i>	Water stains on whole plant and died	4
<i>M. bicolor</i>	Only few leaves were green, putrescence and sear on most leaves	4
<i>C. zebrina</i>	Only 1 leaf blade was green, completely browning and died on other leaves	4

Table 3 FDH concentration per day(mg · m⁻³) in the chamber

Species	Day after being treated						
	1 d	2 d	3 d	4 d	5 d	6 d	7 d
<i>N. nidus</i> cv. Volulum	14.74	12.00	5.96	1.64	0.32	0.00	0.00
<i>C. lubbersiana</i>	13.40	12.45	3.54	1.27	0.28	0.00	0.00
<i>C. ornata</i>	14.09	11.56	1.94	0.36	0.07	0.04	0.01
<i>C. setosa</i>	14.87	13.40	5.63	2.79	0.98	0.12	0.00
<i>C. freddy</i>	14.09	6.07	1.47	0.14	0.00	0.00	0.00
<i>C. roseo-picta</i>	14.09	7.45	2.56	0.35	0.01	0.01	0.01
<i>P. bi furcatum</i>	13.40	4.73	1.23	0.32	0.11	0.07	0.07
<i>C. makoyana</i>	13.40	12.70	5.09	1.89	0.42	0.00	0.03
<i>M. punctatum</i>	14.09	1.11	0.23	0.05	0.00	0.00	0.00
<i>C. crocata</i>	14.74	10.32	4.62	2.99	0.56	0.26	0.04
<i>C. rotundi folia</i>	14.87	8.78	1.94	0.36	0.04	0.00	0.01
<i>N. nidus</i>	14.09	8.87	4.54	1.03	0.18	0.00	0.01
<i>P. fauriei</i>	13.40	7.44	4.66	2.47	1.29	0.47	0.04
<i>P. ensiformis</i> cv. Victoriae	14.87	7.95	6.24	4.30	2.65	1.09	0.26
<i>P. cretica</i> cv. Albolineata	14.09	3.85	1.94	0.62	0.26	0.05	0.03
<i>N. cordi folia</i>	13.40	13.40	5.15	2.23	0.12	0.12	0.05
<i>C. parasiticus</i>	14.74	12.61	7.34	3.30	2.12	0.48	0.08
<i>B. orientale</i>	14.09	8.12	2.60	1.34	0.11	0.08	0.04
<i>M. bicolor</i>	14.09	9.34	1.60	0.07	0.01	0.00	0.00
<i>C. zebrina</i>	14.74	7.29	0.07	0.04	0.04	0.04	0.04
CK	14.87	13.40	13.40	9.26	4.62	2.81	1.11

(right in the frame) subjected to FDH pollution are shown in Fig. 1. The concentration of formaldehyde in the chamber was decreased fast in the first three days. The fastest purification of FDH was found in species such as *C. zebrina*, *M. punctatum*, and the slowest was found in species such as *C. parasiticus*, *P. ensiformis* cv. Victoriae, *N. nidus* cv. Volulum, *C. setosa*, and

N. cordi folia (Table 3).

2. 2 Absorption of FDH by potted plants

As shown in Table 4, the top five plants which had high FDH absorption per dry weight were *N. nidus*, *C. rotundi folia*, *P. cretica* cv. Albolineata, *P. bi furcatum*, and *C. ornata*. The hurt response of potted plants to FDH pollution was divided into 10 scores

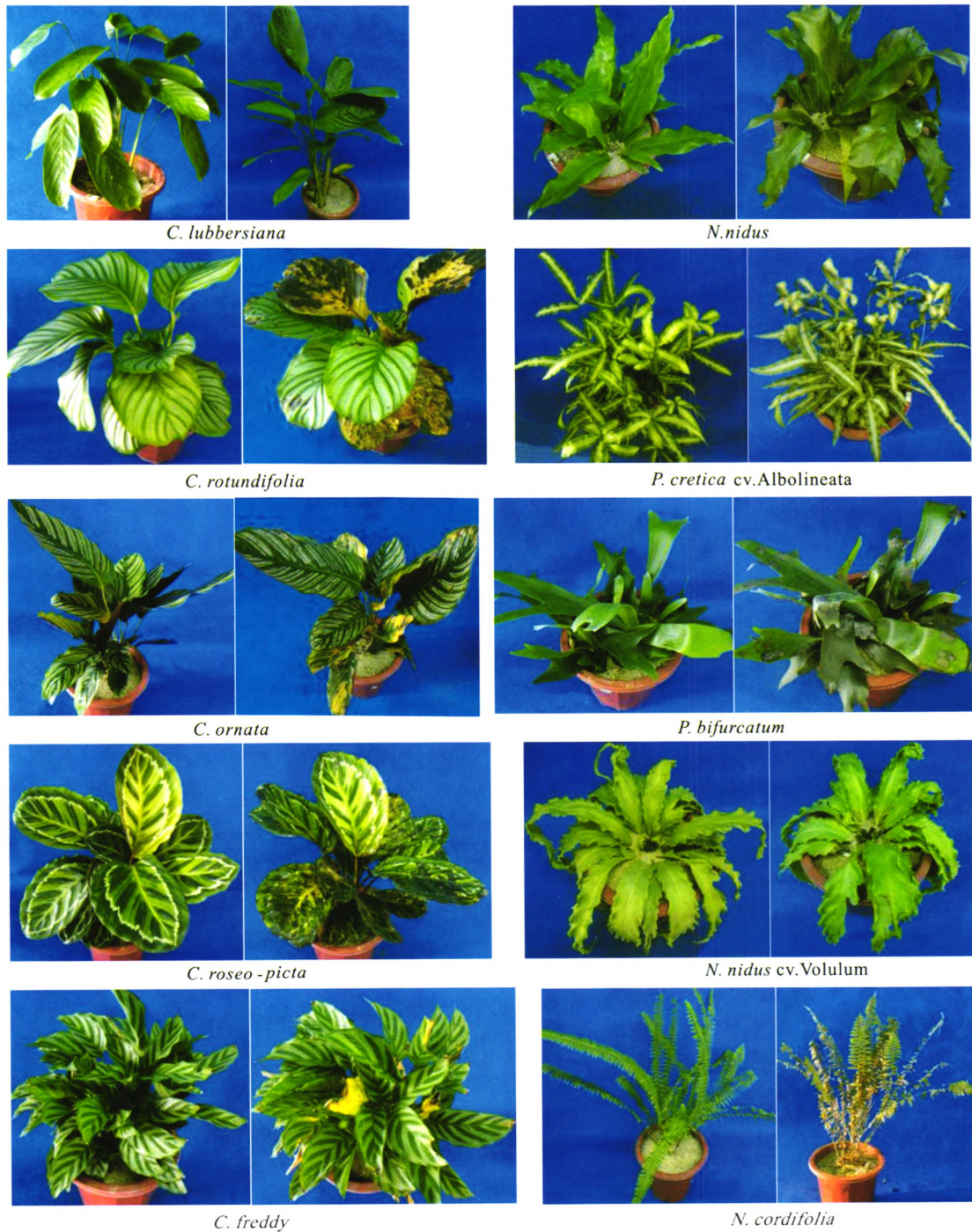


Fig. 1 Photos of some potted plants before (left in the frame) and after (right in the frame) subjected to FDH pollution

according to (Zhou *et al.*, 2011). The score of the FDH absorption per dry mass was made an adjustments, that was, used the 4th day's concentration substituted for the seventh day's. The integrated evaluation on the FDH purification effect of potted plants was shown in Table 5. The top eight plants for FDH purification effect according to their integrated evaluation were listed in Table 5. *N. nidus*,

C. rotundifolia, *P. cretica* cv. Albolineata, *C. ornata*, *P. bifurcatum*, *N. nidus* cv. Volulum, *C. roseo-picta*, and *C. freddy* for the integrated score was surpass 3.60 respectively.

2.3 Changes in total chlorophyll concentration and cell membrane permeability

According to changes in cell membrane permeability (CMP), plants were divided into three groups. The

Table 4 The FDH absorption content and absorption content per dry material of potted plants

Species	Plant DM (g)	Decr in FDH (mg · m ⁻³)	FDH abso (g)	FDHAB per DM (mg · g ⁻¹)
<i>N. nidus</i> cv. Volulum	10.53	13.10	2.84	0.27
<i>C. lubbersiana</i>	21.60	12.12	1.86	0.09
<i>C. ornata</i>	9.94	13.73	3.47	0.35
<i>C. setosa</i>	27.10	12.08	1.82	0.07
<i>C. freddy</i>	13.09	13.96	3.70	0.28
<i>C. roseo-picta</i>	10.86	13.75	3.49	0.32
<i>P. bifurcatum</i>	7.29	13.08	2.82	0.39
<i>C. makoyana</i>	12.81	11.51	1.25	0.10
<i>M. punctatum</i>	18.01	14.04	3.78	0.21
<i>C. crocata</i>	11.59	11.75	1.49	0.13
<i>C. rotundi folia</i>	8.82	14.51	4.25	0.48
<i>N. nidus</i>	4.66	13.06	2.80	0.60
<i>P. fauriei</i>	5.95	10.93	0.67	0.11
<i>P. ensiformis</i> cv. Victoriae	11.48	10.57	0.31	0.03
<i>P. cretica</i> cv. Albolineata	7.00	13.48	3.22	0.46
<i>N. cordi folia</i>	7.92	11.17	0.91	0.11
<i>C. parasiticus</i>	10.15	11.44	1.18	0.12
<i>B. orientale</i>	7.75	12.75	2.49	0.32
<i>M. bicolor</i>	11.58	14.03	3.77	0.33
<i>C. zebrina</i>	13.56	14.70	4.44	0.33

Plant DM, plant dry mass; Decr in FDH, decrease in FDH; FDH abso, actual FDH absorption; FDHAB per DM, actual FDH absorption per dry mass.

Table 5 Integrated evaluation for the purification effects of potted plants on FDH pollution

Species	HR	AF	IS	SO
<i>N. nidus</i> cv. Volulum	8.9	2.7	3.94	6
<i>C. lubbersiana</i>	8.9	0.9	2.50	13
<i>C. ornata</i>	6.9	3.5	4.18	4
<i>C. setosa</i>	6.9	0.7	1.94	15
<i>C. freddy</i>	6.8	2.8	3.60	8
<i>C. roseo-picta</i>	6.8	3.2	3.92	7
<i>P. bifurcatum</i>	4.9	3.9	4.10	5
<i>C. makoyana</i>	4.9	1.0	1.78	16
<i>M. punctatum</i>	4.9	2.1	2.66	12
<i>C. crocata</i>	4.9	1.3	2.02	14
<i>C. rotundi folia</i>	4.9	4.8	4.82	2
<i>N. nidus</i>	2.6	6.0	5.32	1
<i>P. fauriei</i>	2.7	1.1	1.42	18
<i>P. ensiformis</i> cv. Victoriae	2.9	0.3	0.82	20
<i>P. cretica</i> cv. Albolineata	2.8	4.6	4.24	3
<i>N. cordi folia</i>	2.9	1.1	1.46	17
<i>C. parasiticus</i>	1.1	1.2	1.18	19
<i>B. orientale</i>	1.9	3.2	2.94	11
<i>M. bicolor</i>	2.1	3.3	3.06	9
<i>C. zebrina</i>	2.0	3.3	3.04	10

HR, hurt response; AF, absorption fraction; IS, integrated score; SO, sort order.

first group, with change of more than 10%, included 11 species such as *P. ensiformis* cv. Victoriae, *P. fauriei*, *P. cretica* cv. Albolineata, *C. parasitic-*

Table 6 Changes in the total chlorophyll contents and cell membrane permeability of potted plants

Species	Change in chlorophyll (g · kg ⁻¹ FW)	CMP (%)
<i>N. nidus</i> cv. Volulum	0.07	10.49
<i>C. lubbersiana</i>	-0.27	3.28
<i>C. ornata</i>	-0.14	9.31
<i>C. setosa</i>	-0.31	15.51
<i>C. freddy</i>	0.09	5.16
<i>C. roseo-picta</i>	1.63	6.20
<i>P. bifurcatum</i>	0.72	6.08
<i>C. makoyana</i>	0.29	16.64
<i>M. punctatum</i>	1.01	10.57
<i>C. crocata</i>	0.89	8.13
<i>C. rotundi folia</i>	0.73	4.21
<i>N. nidus</i>	0.68	11.97
<i>P. fauriei</i>	1.07	19.46
<i>P. ensiformis</i> cv. Victoriae	0.48	21.82
<i>P. cretica</i> cv. Albolineata	2.13	18.96
<i>N. cordi folia</i>	0.99	14.73
<i>C. parasiticus</i>	-3.28	16.94
<i>B. orientale</i>	-1.52	4.07
<i>M. bicolor</i>	-1.13	16.54
<i>C. zebrina</i>	0.11	7.75

CMP, cell membrane permeability.

us *C. makoyana*, *M. bicolor*, *C. ornata*, *N. cordi folia*, *N. nidus*, *M. punctatum*, and *N. nidus* cv. Volulum. The second group with CMP change between 5% and 10% included 6 species such as *C. ornata*, *C. crocata*, *C. zebrina*, *C. roseo-picta*, *P. bifurcatum*, and *C. freddy*. The third group with CMP changes less than 5% included 3 species such as *C. rotundi folia*, *B. orientale*, and *C. lubbersiana*. However, there were no obvious relations of the FDH absorption ability with both the changes in total chlorophyll concentration, and the changes in cell membrane permeability. It maybe suggested that the lower the change in cell membrane permeability was, the stronger the FDH absorption ability was.

3 Discussion

Currently, it seems a bit confusing in ranking of the ability of plants to absorb and purify FDH. We suggested top 8 species in purification of FDH as *N. nidus*, *C. rotundi folia*, *P. cretica* cv. Albolineata, *C. ornata*, *P. bifurcatum*, *N. nidus* cv. Volulum, *C. roseo-picta*, and *C. freddy*. Surprisingly, there were 4 plants from Pteridophytes and 4 plants from Marantaceae in

the top plants, outstanding for their low hurt response and high absorption ability. We could not compare our results with other researchers (Zhou *et al.*, 2006; Li 2006; Wang *et al.*, 2007; Huang *et al.*, 2008; Cao *et al.*, 2009; Xiong & Su, 2009; Tian *et al.*, 2011). Although they tried to give the ranking of FDH purification ability, the number of tested potted plants were both few and insystemic, or there were some problems in their experiment designs. For example, they wrapped or sealed the pot, media and the bottom of the tested plants with plastic bags or films (Zhou, *et al.*, 2006; Achkor *et al.*, 2003; Song *et al.*, 2007), without enough number of plants, in turn, without enough tillers or shoots, which could not ensure high FDH absorption. There may be three ways for potted plants to react to FDH air pollution. The first with high absorption but weak resistance to FDH damage, showed obvious hurt morphology, for example, in plants such as *N. cordifolia*, *P. fauriei*, *C. parasiticus* and *P. ensiformis* cv. Victoriae. The second showed weak absorption but strong resistance with normal morphology by taking avoidance strategy to protect itself. Plants such as *C. lubbersiana* and *C. ornata* belong to this group. The third showed absorption and transforming ability with more or less hurt responses, included plants such as *C. rotundifolia*, *C. setosa*, and *C. roseo-picta*. It was perplexed that some plants had higher absorption ability for FDH, but had lower cell membrane permeability, vice versa. This mechanism may be need further studies in future.

4 Conclusions

In conclusion, 8 species of the indoor potted plants which could be recommendable to be used for formaldehyde purification were *N. nidus*, *C. rotundifolia*, *P. cretica* cv. Albolineata, *C. ornata*, *P. bifurcatum*, *N. nidus* cv. Volulum, *C. roseo-picta*, and *C. freddy*.

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References:

- Achkor H, Díaz M, Fernández MR, *et al.* 2003. Enhanced formaldehyde detoxification by overexpression of glutathione-dependent formaldehyde dehydrogenase from *Arabidopsis*[J]. *Plant Physiology*, **132**(8):2 248—2 255
- Cao SJ, Pan BH, Tian YC, *et al.* 2009. Comparison of ability of absorbing formaldehyde among 6 species of indoor ornamentals [J]. *Ecol Environ Sci*, **18**(5):1 798—1 801(In Chinese)
- Huang AK, Li N, Tang GG. 2008. Capability to absorb high concentration benzene and formaldehyde of four indoor potted plants [J]. *J Environ Health*, **25**(12):1 078—1 080(In Chinese)
- Huang XL, Chen RZ, Zhang BZ. 1990. Seed Physiology Laboratory Manual[M]. Beijing: Agriculture Press:122—124(In Chinese)
- Li QJ. 2006. The ornamental absorb formaldehyde in the room Northeast Forestry University[J]. *Master Thesis*, 15—19(In Chinese)
- Song JE, Kim YS, Sohn JY. 2007. A study on reduction of volatile organic compounds in Summer by various plants[J]. *J S Chin Univ Technol : Nat Sci Edit*, **35**(Suppl):219—222
- Tian YC, Pan BH, Cao SJ. 2011. Research of eight kinds of indoor foliage plants to formaldehyde purification[J]. *Northern Hort*, **2**:82—84(In Chinese)
- Wang YL, Yang ZD, Deng RY, *et al.* 2007. Study on response of several garden plants to formaldehyde pollution[J]. *Guangxi Sci*, **14**(2):163—166(In Chinese)
- Wang XQ. 2006. Principles and Techniques for Plant Physiology and Biochemical Experiment[M]. 2nd ed. Beijing: High Education Press:134—136(In Chinese)
- Wolverton BC. 1997. How to Grow Fresh Air[M]. New York: Penguin Books:18—19
- Wolverton BC, Donald RC, Mesick HH. 1985. Foliage plants for the indoor removal of the primary combustion gases carbon monoxide and nitrogen oxides[J]. *J Mississippi Acad Sci*, **30**:1—8
- Wu P. 2006. Study on the Indoor Pollution Air - Formaldehyde Purification Ability with Several Plants[M]. Nanjing Forestry University, Nanjing. Master thesis, :12—16(In Chinese)
- Xiong Y, Su ZG. 2009. A research on the ability of absorbing formaldehyde among five species of indoor ornamentals[J]. *Environ Sci Manag*, **34**(1):45—47(In Chinese)
- Yu GL, Tang HY. 2005. Investigation and analysis of the indoor environment of the new building in Guangzhou city[J]. *Chin J Health Insp*, **15**(3):350—364(In Chinese)
- Yu Y, Yang Y, Liu XD, *et al.* 2007. Probe on the resistant ability of plants to formaldehyde[J]. *Sci Technol Inf*, **16**:40, 59(In Chinese)
- Zhou JH, Qin FF, Su J, *et al.* 2011. Purification of formaldehyde-polluted air by indoor plants of Araceae, Agavaceae and Liliaceae [J]. *J Food Agric & Environ*, **9**(3, 4):1 012—1 018
- Zhou XJ, Liang SY, Jin YJ, *et al.* 2006. The comparison of ability of absorbing formaldehyde among 13 species of indoor ornamentals[J]. *Chin Agric Sci Bull*, **22**(12):229—231(In Chinese)

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参考文献(17条)

- [Aehkor H;Díaz M;Fernández MR Enhanced formaldehyde detoxification by overexpression of glutathione-dependent formaldehyde dehydrogenase from Arabidopsis 2003\(08\)](#)
- [曹受金, 潘百红, 田英翠, 王贵武 6种观赏植物吸收甲醛能力比较研究\[期刊论文\]-生态环境学报 2009\(5\)](#)
- [黄爱葵, 李楠, 汤庚国 四种室内盆栽植物对高浓度苯和甲醛的吸收特性\[期刊论文\]-环境与健康杂志 2008\(12\)](#)
- [Huang XL;Chen RZ;Zhang BZ Seed Physiology Laboratory Manual 1990](#)
- [Li QJ The ornamental absorb formaldehyde in the room 2006](#)
- [Song JE;Kim YS;Sohn JY A study on reduction of volatile organic compounds in Summer by various plants 2007\(Suppl\)](#)
- [Tian YC;Pan BH;Cao SJ Research of eight kinds of indoor foliage plants to formaldehyde purification \(In Chinese\) 2011](#)
- [王利英, 杨振德, 邓荣艳, 覃寿艺 几种园林植物对甲醛污染的反应研究\[期刊论文\]-广西科学 2007\(2\)](#)
- [Wang XQ Principles and Techniques for Plant Physiology and Biochemical Experiment 2006](#)
- [Wolverton BC How to Grow Fresh Air 1997](#)
- [Wolverton BC;Donald RC;Mesick HH Foliage plants for the indoor removal of the primary combustion gases carbon monoxide and nitrogen oxides 1985](#)
- [Wu P Study on the Indoor Pollution Air-Formaldehyde Purification Ability with Several Plants 2006](#)
- [熊纓, 苏志刚 五种常见装饰植物对甲醛的吸收能力比较研究\[期刊论文\]-环境科学与管理 2009\(1\)](#)
- [Yu GL;Tang HY Investigation and analysis of the indoor environment of the new building in Guangzhou city \(In Chinese\) 2005\(03\)](#)
- [Yu Y;Yang Y;Liu XD Probe on the resistant ability of plants to formaldehyde 2007](#)
- [Zhou JH;Qin FF;Su J Purification of formaldehydepolluted air by indoor plants of Araceae, Agavaceae and Liliaceae 2011\(3,4\)](#)
- [周晓晶, 梁双燕, 金幼菊, 王骏, 赵惠恩 13种常用室内观赏植物对甲醛净化效果\[期刊论文\]-中国农学通报 2006\(12\)](#)

引证文献(2条)

- [欧坚泉, 周俊辉, 陈水渐, 岳保超 几种盆栽植物对甲醛的净化作用\[期刊论文\]-北方园艺 2012\(22\)](#)
- [何勤勤, 周俊辉 盆栽植物对室内甲醛空气污染的净化研究进展\[期刊论文\]-江西农业学报 2014\(02\)](#)

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